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Tailoring Nanodiamonds for High-Contrast EPR Imaging: Size, Surface Properties, and Spectroscopic Performance

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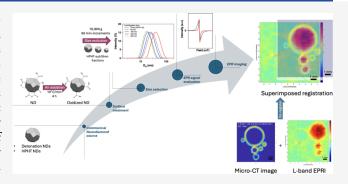
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s ABSTRACT: Electron paramagnetic resonance (EPR) spectros-6 copy is a tool that provides sensitive detection of uncoupled 7 electron spins for a variety of applications. This technique enables 8 the specific detection and quantification of radical species while 9 also being able of generating high-contrast, background-free 10 images. However, the EPR labeling and imaging techniques 11 encounter limitations mainly due to the instability of organic 12 radicals from organic probes, which can influence the reliability 13 and scope of the experiment. In that context, the use of 14 nanodiamonds (NDs) in EPR may be a promising route for 15 understanding their unique properties and potential biomedical 16 applications. The ability to perform EPR imaging in combination



with the stable intrinsic properties of paramagnetic centers within these particles raises the possibility of extending nanodiamond18 based imaging capabilities. Herein, we present a preliminary demonstration of a practical spectroscopy and imaging application using
19 nanosized diamond particles (<18 nm) for electron paramagnetic resonance imaging (EPRI). The discretization of two different
20 nanodiamond production sources among the most studied NDs (HPHT or detonation) allows further characterization of their
21 physicochemical properties. In addition, we have investigated variations in the physicochemical properties of nanodiamonds,
22 including size effects and surface treatments. Finally, we provide experimental evidence of the conditions required for optimal
23 spectroscopic and imaging resolution (R < 1 mm) as well as achievable EPR sensitivity.

24 INTRODUCTION

25 Electron paramagnetic resonance (EPR), as a spectroscopic 26 technique, is specific to the resonance of paramagnetic species 27 such as (in)organic radicals, complexes, crystallographic 28 defects, colored centers, or transition metal ions when 29 subjected to a magnetic field. Many concepts in electron 30 spin (or paramagnetic) resonance (ESR or EPR) spectroscopy 31 are related to similar notions in nuclear magnetic resonance 32 (NMR) spectroscopy; like what NMR brings to MRI 33 modality, EPR can enable EPR imaging (EPRI) using an 34 additional magnetic field gradient in a set of different 35 orientations around the sample. High-resolution images using 36 a noninvasive imaging technique enable the specific detection 37 and quantification of paramagnetic species with optimal 38 resolution. The choice of the probe depends on the specific 39 application and the physicochemical properties of the sample 40 under study. Overall, the versatility and sensitivity of EPR 41 probes make them essential tools for a wide range of 42 applications in medicine, industry, and the environment. The 43 best-known probes and their applications include (i) nitroxide 44 species for studying biological systems (i.e., superoxide spin 45 trapping), 3,4 (ii) triarylmethyl radicals (TAMs) for oximetry 46 (i.e., to measure oxygen concentration in biological samples for

in-depth assessment of tissue oxygenation, metabolism, and 47 redox evaluations), 5,6 and (iii) site-directed spin labeled 48 (SDSL) nitroxides to evaluate structural and dynamic 49 properties of organic systems, such as transmembrane 50 transport and transcription mechanisms or as Cu(II) spin 51 labels. The However, the stability of paramagnetic organic 52 compounds can be influenced by a number of factors such as 53 pH, temperature, or redox environment, influencing the 54 reliability of the experiment. The probes, specifically tailored 55 for paramagnetic labeling, can be assessed through EPR studies 56 to obtain valuable information.

While we focus on the capabilities of EPR spectroscopy to 58 analyze paramagnetic materials, particular attention is paid to 59 nanodiamonds (NDs), whose unique properties are attracting 60 growing interest in a variety of scientific domains. Integrating 61 the EPR properties of NDs into the EPR imaging modality 62

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63 enables in-depth characterization of these nanoplatforms, 64 opening up new perspectives in applications ranging from 65 nanomedicine to materials science. Indeed, nanodiamonds 66 have emerged as promising carbon-based nanomaterials for a 67 range of applications due to their unique advantages, 68 particularly in the biological, medical, and imaging fields. 10-12 69 In these fields, the nitrogen-vacancy (NV) centers embedded 70 in the core of the NDs are exploited for their ultrastable 71 fluorescence properties, while some of the fluorescent probes 72 are limited either by their photostability or cytotoxicity. 13-15 73 Other interests include the use of crude NDs as noncytotoxic 74 substrates for gene and drug delivery, 11,16 as implants, 17 in 75 tissue engineering or as powerful theranostic substrates for the 76 targeted delivery of vaccines, 18 as chemotherapy agents, 16,19 in 77 immunotherapeutics, as Gd- and Mn-doped (grafted) DND 78 MRI contrast agents, ^{20–24} or for hyperpolarization MRI^{25–28} using ND intrinsic impurities.

Apart from optical fluorescence, NMR, or hyperpolarization purposes, options for noninvasive in vitro and in vivo imaging 82 of nanodiamonds are very limited. Three main advantages of 83 using NDs in EPRI can be noted: The first one is closely linked 84 to the relative stability of the radical-like paramagnetic centers 85 (in the bulk and the core of the particle) compared with 86 conventional organic radicals (i.e., easy reduction through 87 metabolic pathways).²⁹ The second results from the nano-88 particle unique characteristics; nanoparticles provide a versatile 89 platform that can be used in a variety of biomedical and 90 technological applications regarding their distinct physico-91 chemical properties (enhanced surface area, properties 92 tailoring, biocompatibility, tunable surface chemistry, and 93 surface charge or versatility). 30 The third results from the 94 EPR signal; the EPR signal of NDs typically exhibits a narrow 95 line indicating well-defined electron spin states resulting in a 96 high signal-to-noise ratio (as a function of line width). In 97 addition, the EPR spectrum may display other specific features 98 corresponding to (structural) defects or impurities in the diamond lattice depending on the material studied (nano-100 diamond source, size, surface graphitization, and crystallinity). Two primary methods for producing nanoscale diamond 102 particles have been introduced based on their production 103 procedures: (i) a detonation-based synthesis where the carbon 104 source is derived from the explosives or (ii) a mechanical 105 grinding of high-quality HPHT (high-pressure high-temper-106 ature) diamond microcrystals into nanometric particles. 107 Particles obtained by detonation are rather small (<10 nm) 108 and relatively monodisperse but exhibit poor crystalline 109 quality, whereas those obtained by fragmentation of bulk 110 material exhibit superior crystallinity, albeit with a broader size 111 distribution (from hundreds of micrometers to a few 112 nanometers).^{31,32} Depending on the nanodiamond synthesis 113 procedure, their properties (size, structure, surface composi-114 tion, EPR or optical luminescence properties, and crystallinity) 115 can be significantly impacted. For example, there are many 116 more defects in ND_{DET} compared to ND_{HPHT} as a result of the 117 uncontrolled explosion event. These include vacancies, 118 dislocations, domain boundaries, and amorphous carbon 119 impurities, mainly on the surface and within the interface 120 layer (i.e., dangling bonds). 31,32 In contrast, ND_{HPHT} defects 121 consist of vacancies, dislocations, or stacking faults in a 122 crystalline lattice. Furthermore, their surface properties are 123 very different, with ND_{DET} having a complex surface with a

124 higher density of defects, amorphous and graphitic carbons,

and various functional groups, leading to aggregation due to 125 weak electrostatic interactions. 31,32

Ultimately, the choice between both types depends on 127 specific requirements, such as particle size distribution, purity, 128 surface chemistry, or fluorescence stability. In general, the 129 small-sized detonation nanodiamond is the typical choice for 130 MRI purposes, while larger-sized HPHT particles are suitable 131 in tissue engineering or luminescence purpose. 33

Here, we present the physicochemical property variations 133 from production source, size, and surface treatments effects 134 and EPR investigation. By means of mathematical develop- 135 ments, imaging optimizations, and a reliable nanodiamond 136 strategy, we demonstrate EPRI capabilities using this nano- 137 material. To our knowledge, this is the first experimental 138 evidence of EPR imaging using NDs.

140

MATERIALS AND METHODS

Materials. The monocrystalline synthetic microdiamond powder 141 of HPHT origin (Microdiamant MSY 0-0.03) was supplied by Pureon 142 AG (Lengwil, Switzerland). Primary particles of the detonation 143 nanodiamond were manufactured by NanoAmando (NanoCarbon 144 Research Institute Ltd., Nagano, Japan). Branched poly- 145 (ethylenimine) (MW = 1200 Da, ≥99%) and ICP zirconium plasma 146 standard were supplied by ThermoFischer Scientific (Merelbeke, 147 Belgium). ICP multielement standard solution was supplied by Merck 148 (Hoeilaart, Belgium). All chemicals used were of standard purity 149 grade. Stirred cells and membranes (MWCO = 100 kDa, RC) for 150 ultrafiltration were purchased from Metrohm (Antwerpen, Belgium). 151

Preparation of Nanodiamond Suspensions. Nanodiamond 152 suspensions were prepared by dispersing diamond powders (200 mg) 153 either used as-received ("asrec") or thermally oxidized ("ox") in 154 deionized (DI) water. To ensure a good colloidal dispersion, particles 155 were sonicated in DI water for 90 min (10 min steps) using an 156 ultrasound probe (Hielscher, 200 W, 24 kHz) with an amplitude of 157 70% (duty cycle: 1/2) and slightly centrifuged (670g, 5 min) to 158 exclude larger aggregates. Nanoparticle density was obtained gravi- 159 metrically after freeze-drying 1 mL of the suspension followed by a 160 drying step at 80 °C for 24 h. If necessary, nanodiamonds were 161 concentrated by a stirred ultrafiltration cell device (MWCO = 100 162 kDa) to reach a particle concentration of 20 mg mL⁻¹.

Oxidative Treatment by Air Annealing. A ceramic crucible 164 filled with crude freeze-dried diamond powder was heated in a muffle 165 furnace (heating rate: 10 °C min⁻¹) in air at 400 °C (ox-1), 450 °C 166 (ox-2), 480 °C (ox-3), or 550 °C (ox-4) for 4 h (Nabertherm GmbH, 167 Lilienthal, Germany) to remove the graphitic shell and modify 168 oxidation-sensitive chemical groups. The conditions of oxidative air 169 treatment by air annealing were studied by thermal analysis (TGA) 170 and carried out in accordance with published procedures with some 171 modifications. The air-annealed (oxidized) NDs were then 172 dispersed in DI water as previously described to obtain a gray/ 173 black nanodispersion (20 mg mL⁻¹).

Size Exclusion Process. Sub-17 nm $\mathrm{ND}_{\mathrm{HPHT}}$ fractions were 175 obtained by successive high-speed centrifugation cycles on $\mathrm{ND}_{\mathrm{HPHT}}^{\mathrm{assec}}$ 176 (5 mg mL $^{-1}$). Between each centrifugal treatment (IEC CL31R 177 Multispeed Centrifuge, 16,800g; 0 < n < 7 × 60 min steps), the 178 supernatant was carefully isolated, characterized, and treated again. 179 Their notations throughout the article are written as follows: $\mathrm{ND}_{\mathrm{HPHT}}^{1}$ 180 isolated after a centrifugation cycle of 1 × 60 min; $\mathrm{ND}_{\mathrm{HPHT}}^{2}$ sample 181 isolated after 2 × 60 min, etc. This procedure was not applied to 182 detonation nanodiamonds due to a lack of colloidal stability resulting 183 from their aggregation state.

Physicochemical Characterization Techniques. The hydro- 185 dynamic particle size and zeta potential were determined by photon 186 correlation spectroscopy (PCS) using a Zetasizer Nano ZS particle 187 size analyzer (Malvern Instruments, He—Ne laser, 633 nm, scattering 188 angle 173°) (Worcestershire, UK) on diluted suspensions (1 mg 189 mL⁻¹ in DI water, 25 °C). The refractive index of bulk diamond (2.4 190 at 635 nm) was used to evaluate the intensity and number-weighted 191

192 distributions. PCS measurements are presented as mean values 193 (hydrodynamic $D_{\rm H}^{\rm PCS}$ or number diameters ($D_{\#}^{\rm PCS}$) and zeta 194 potential (ζ)) \pm standard deviation. The evolution of the zeta 195 potential was investigated in relation to pH changes; the pH of each 196 nanodiamond sample was adjusted using diluted HNO₃ or TMAOH 197 solution.

TEM experiments were performed with a Fei Tecnai 10 electron 198 199 microscope (Oregon, USA) operating at an accelerating voltage of 80 200 kV under a vacuum to obtain detailed information on the morphology 201 and the size distribution of a sample. Each specimen was prepared 202 following a procedure slightly adapted from Rehor and Cigler.³⁶ 203 Briefly, 300 mesh carbon-coated Formvar grids (Ted Pella Inc., 204 California, USA) were pretreated using an UV/ozonizing chamber 205 (AppliTek Scientific Instruments, Nazareth, Belgium) for 15 min. A 206 mixture droplet (4 μ L) of a branched ethylenimine polymer solution 207 (MW = 1,2 kDa, 0.1 mg mL⁻¹) and a 0.1 mg mL⁻¹ colloidal 208 suspension was then placed on the grid, allowing the liquid to dry in 209 air at room temperature. The statistical particle size distribution was 210 obtained by examining multiple images of each sample using the 211 iTEM software (Münster, Germany) using the Feret diameter as the 212 size expression due to the irregular shape of the studied ND_{HPHT} 213 particle. By measuring the diameter size of 400 to 500 counted 214 particles for each sample, the mean equivalent median diameter 215 (D^{TEM}) , the polydispersity index $(\text{PDI}^{\text{TEM}})$, and a standard deviation 216 (SD) from the corresponding particle suspension were calculated.³⁷ A 217 size histogram showing the number-weighted distribution was 218 obtained, and the resulting histograms were fitted with a log-normal 219 function.

Thermogravimetric analyses (TGAs) were performed on a TA 221 Q500 device (TA Instruments, New Castle, USA). The mass loss of 222 predried HPHT and DET samples monitored under air was over the 223 process for thermal stability investigation. After an isotherm at 120 °C 224 for 10 min (heating rate: 10 °C min $^{-1}$), the temperature was 225 increased from 120 to 800 °C at a heating rate of 10 °C min $^{-1}$ under 226 air flow (25 mL min $^{-1}$), and the temperature was maintained for 5 227 min.

The Boehm titration method was evaluated on ND $_{
m DPT}^{
m arresc}$, ND $_{
m ND}^{
m CPT}$, ND $_{
m CPT}^{
m ND}$, 229 and ND $_{
m HPHT}^{
m arresc}$ nanoparticles (10 mg mL $^{-1}$) under conductometric 230 detection (SI Analytics, Lab 945, Xylem) to quantify the oxygenated 231 functional groups present on the particle surface according to a 232 published procedure. ³⁸

233 Infrared spectra (ATR) of dried powders were recorded on a 234 PerkinElmer FTIR Spectrum 100 spectrometer (Manchester, UK) 235 with an average of four scans and a resolution of 4 cm^{-1} .

Raman measurements were performed on an Xplora Raman 237 spectrometer (Horiba) under green wavelength laser excitation (532 238 nm, 0.79 mW) equipped with a 1800 tr/mm grating. Prior to their 239 characterizations, samples were dispersed in DI water and dispensed 240 onto a Si substrate by drop-casting for analysis. Spectra have been 241 recorded with a 10 s acquisition time, repeated 10 times. To ensure 242 the representativeness of the spectra presented here, three different 243 areas of the samples were systematically probed.

The surface chemistry of the nanodiamond samples was 245 characterized by X-ray photoelectron spectroscopy (XPS) using an 246 XPS Versa Probe Phi 5000 spectrometer (Chanhassen, USA). The X-247 ray source was a monochromatized Al K α line ($E_{\rm b}=1486.7~{\rm eV}$). 248 Measurement of the deconvoluted C Is and O Is orbital peak energies 249 was carried out using a linear baseline and Gaussian line curves of 250 variable widths.

The elemental composition of the nanodiamonds was determined by high-resolution TEM energy-dispersive X-ray spectroscopy (HRTEM-EDX) scanning with an average of three measurements for the qualitative measurements (TEM grids: Formvar/carbon supported copper grids). Prior to quantitative analysis of impurities, the as-received detonation and HPHT powders (250.0 \pm 0.1 mg) were calcinated in a muffle furnace (heating rate: 10 °C min⁻¹; 650 cs °C for 3 h). The resulting yellow/light brown residues (6.3–3.4 mg, 259 respectively) were then quantitatively transferred into Teflon vessels for microwave acid digestion (Milestone MLS 1200 Mega, Analis, 261 Belgium). Levels of total uncalcinated impurities of 2.52 and 1.36% w/w

for DET and HPHT powders, respectively, were measured gravi- 262 metrically, corresponding to 95-98% of the major carbon element in 263 NDs. The volumes of the resulting acid-treated samples were then 264 adjusted to 25 mL using DI water. The impurity (i.e., Ag, Al, Zn, Fe, 265 Cr, Cu, Zr, and Mn) content was quantified by inductively coupled 266 plasma-atomic emission spectroscopy (ICP-AES) under argon plasma 267 on a Varian Liberty Series II instrument (Varian Inc., Palo Alto, 268 USA). Calibration curves (26 > 269) using different dilutions of an 269 ICP multielement standard solution were constructed to ensure the 270 proper quantification of each metallic impurity.

EPR Spectroscopy and Imaging. EPR Spectroscopy. X-band 272 electron paramagnetic resonance spectroscopy was used to identify 273 the paramagnetic contributions to the overall EPR spectrum and to 274 obtain quantitative information about paramagnetic centers. Samples 275 (20 mg mL⁻¹ in DI water) were analyzed at room temperature on a 276 Bruker EMX nano (Bruker, Rheinstetten, Germany). The following 277 spectroscopic parameters were used: 343.5 mT center field, 278 microwave frequency of ~9.8 GHz, magnetic field modulation 279 amplitude of 0.1 mT, and frequency of 100 kHz and presented as 280 the derivative of the microwave absorption over the magnetic field. In 281 the specific case of nanodiamonds, their EPR spectra typically present 282 a single line characterized by a narrow or large resonant shape, 283 depending on the electron spin states. Therefore, the type of 284 (isotropic) broadening applied to a line width (lw) is referred to as 285 lwpp (pp, peak-to-peak) to measure the horizontal distance between 286 the maximum and the minimum of the first-derivative EPR line shape. 287 The g-factor refers to the spectroscopic splitting factor, characterizing 288 the behavior of species in a magnetic field, and is extracted from the 289 minima in the second-derivative EPR spectra.

Spectroscopic Simulation. EPR spectra were simulated using the 291 EasySpin software package (Matlab). 40 292

Low-Frequency EPR Settings. Low microwave spectroscopic EPR 293 data were recorded at 1 GHz (L-band) on a CW EPR E540L 294 spectrometer (Bruker, Rheinstetten, Germany) at room temperature. 295

Phantom Preparation. Nanodiamond suspensions in DI water 296 (filled with $100-200~\mu\text{L}$ at a $10~\text{mg mL}^{-1}$ particle density) were 297 placed in EPR tubes (4, 1, 0.5, 0.4, and 0.3 mm quartz capillary 298 diameter size) inserted into a 23 mm birdcage resonator mounted 299 inside the device for phantom imaging study.

Phantom EPR Imaging Experiments. The EPR imaging was 301 performed using the L-band spectrometer. The continuous wave 302 system was equipped with gradient coils to obtain information about 303 the spatial distribution of free radicals in the nanodiamond samples. 304 Spectrometer parameters were as follows: microwave power: 50 mW; 305 field of view (FOV) diameter: 82.4 mm; sweep width: 8.24 mT; 306 microwave frequency: 1.105 GHz; amplitude modulation: 0.2 mT; 307 and frequency modulation: 50 kHz. Experimental resolutions were 308 evaluated on the basis of images of quartz capillaries (0.3–4 mm 309 diameters) filled with nanodiamond suspensions with repositioning 310 on anatomical images obtained via X-ray microcomputed (micro-CT) 311 tomography.

Image Reconstruction and Mathematical Procedures. EPR 313 image reconstruction was performed using a Total Variation 314 algorithm-based, 41 published and usable online. 42 315

X-ray Microtomographs. Micro-CT imaging was performed using 316 a 1178 X-ray computed scanner (Skyscan, Kontich, Belgium) with the 317 following parameter setting: 65 kV, 615 μ A, and pixel size 83.82932 318 μ m. The spatial resolution of quartz capillary diameters was 319 determined by analyzing the micro-CT images using the Skyscan 320 software suite. The images were co-registered using GIMP.

■ RESULTS AND DISCUSSION

Physicochemical Characterization of NDs (As-Re- 323 ceived ND_{DET} and ND_{HPHT}). For ND_{HPHT}, we used as- 324 received (asrec) commercially available MSY 0-0.03 powders 325 as these are the smallest monocrystalline particles reported. 326 TEM analysis revealed NDs with irregular shapes with sharp 327 edges and a large size distribution (17 nm; PDI TEM: 1.79). The 328 particle size distributions obtained after sonication show 329

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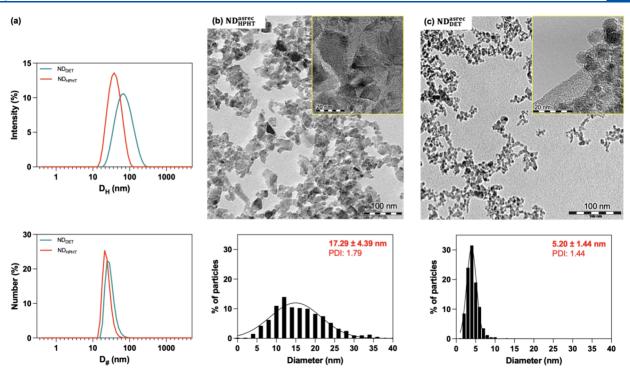


Figure 1. Size measurements. (a) Size distributions obtained by PCS (intensity-weighted (up) and number-weighted (bottom) distributions). TEM images of (b) mean 17 nm ND_{HPHT} and (c) mean 5 nm ND_{DET} (scale bars: 100 nm) and their particle size distributions. The α axis for ND_{HPHT} particle histograms is expressed as a 2 nm increment size diameter for clarity (1 nm increment size for ND_{DET}). Extensions (yellow frames) show high-resolution TEM images (scale bars: 20 nm) obtained by the HRTEM-EDX experiment (as-received HPHT and detonation samples: 200k and 250k× magnification, respectively).

330 narrow, almost monodisperse particle distributions (24 nm in 331 number-weighted distributions; Figure 1a), consistent with $_{
m 332}$ TEM observations. For ND $_{
m DET}$, while small-sized particles were observed by TEM (5.2 nm, PDI^{TEM}: 1.44), a larger 334 average particle size was observed using DLS (63 and 32 nm in 335 intensity- and number-weighted distributions, respectively), 336 suggesting the dispersion of aggregates rather than individual particles. Contrary to ND_{HPHT}, ND_{DET} was characterized by 338 low colloidal stability ($\zeta = -39$ and +42 mV, pH 7.3), 339 especially when the ionic strength of the medium is increased, 340 which could reflect significant differences in its surface chemical composition. IR analysis confirmed this hypothesis 342 by revealing a characteristic band at 1766 cm⁻¹, corresponding 343 to the C=O stretching of carboxylic acid groups, present only 344 on ND_{HPHT} (Figure S2b). This feature is responsible for the 345 negative zeta potential that ensures the colloidal stability. On 346 both types, additional signatures of oxidized terminations, such 347 as alcohols and epoxy groups, are indicated by bands at 1290 348 and 1090 cm⁻¹ corresponding to symmetric and asymmetric 349 C-O stretching modes, respectively. Bands at 3670-2670 and 350 1620 cm⁻¹, associated with the O-H stretching and bending 351 modes, respectively, are also observable. These may originate 352 from alcohol functions or from adsorbed water on the ND 353 surface. Finally, ND_{DET} shows a structure between 2800 and 354 2950 cm⁻¹ corresponding to aliphatic C-H stretching modes 355 due to hydrogen terminations and/or nondiamond phases on 356 the surface.

Using high-frequency (X-band, 9 GHz) EPR acquisitions, 358 we can assess that both ND_{HPHT} and ND_{DET} (10 mg mL⁻¹) 359 samples contain electron spins with a main spectroscopic 360 splitting factor (g-factor) = $g_e = 2.0027 \pm 0.0002$. Among 361 impurities other than carbon, some paramagnetic and

ferromagnetic impurities originating from transition metal 362 ions (Fe³⁺, Cr³⁺, Mn²⁺, Co²⁺, Ni²⁺, or Cu²⁺) can be found in 363 diamond suspensions. HR-TEM energy-dispersive X-ray 364 (EDX) confirmed the presence of Al, Fe, or Zr (Table S1) 365 randomly distributed within the bulk, among other elements, 366 with uniform distribution.^{39,44} As a result of the grinding and ₃₆₇ explosion processes (milling with zirconium oxide micrometer- 368 sized beads; sonic mechanical disintegration of corrosion 369 products from the explosion chamber wall), noncarbon 370 elements were incorporated within the suspension. ICP-AES 371 analysis estimated the bulk composition of ND_{DET} and 372 ND_{HPHT} suspensions. The resulting concentrations normalized 373 per ND suspension are given in Table S2. Among the most 374 notable differences between both samples, one has to mention 375 the high zirconium content detected in the ND_{DET} sample 376 (with around 14 ng per mg ND_{DET} and below the LOD in the 377 ND_{HPHT} sample) as a consequence of its dispersion process 378 (ND_{DET} dispersion is achieved using the zirconia beads- 379 assisted method).

Figure 2 shows the main resonance EPR single lines at 343 $_{381}$ fz mT (extension) for the ND_{HPHT} and ND_{DET} samples, resulting $_{382}$ mainly from structural defects (dangling bonds). For small- $_{383}$ sized nanodiamonds (<70 nm), it can be assumed as a two- $_{384}$ component deconvolution since the EPR line has a larger line $_{385}$ width, with a similar signal resonance $_{g}$ -factor mainly attributed $_{386}$ to the free electron resonance ($_{g}$ = 2.0027) but with different $_{387}$ line width contributions: $_{45}$ (i) a broad spin-1/2 Lorentzian $_{388}$ component assigned to C–C carbon dangling bonds on the $_{389}$ particle surface and in the nanoparticle core and (ii) a narrow $_{390}$ spin-1/2 Lorentzian component attributed to defects in the $_{391}$ diamond lattice. $_{45}$ Due to the relatively high local spin $_{392}$ densities and their close proximity, a main characteristic of a $_{393}$

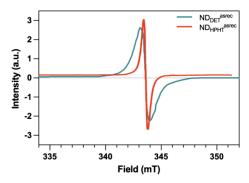


Figure 2. Typical analysis of high-frequency (X-band) EPR profiles of ND $_{\rm DET}^{\rm asrec}$ (green) and ND $_{\rm HPHT}^{\rm asrec}$ (red) suspensions (10 mg mL $^{-1}$) recorded at room temperature (main resonance signal) around the resonant magnetic field of the signal (g=2.0027). Microwave frequency: 9 GHz.

394 nanosized system, these defects are able to mutually interact 395 and, additionally, can interact with nearby P1 centers, 396 increasing the line width. Here, the main differences between

nanodiamonds are the production origin, crystallographic core, 397 internal/surface defects, purification, surface chemistry, 398 agglomeration state, presence of sp^2 -carbon, and material 399 size. The experimental resonance measured on the NDasrec and 400 NDasrec and 400 NDasrec and 400 NDasrec and 401 and 0.38 mT, respectively, at a resonance field $B_0 \sim 343.5$ mT. 402 To understand the surface etching modification, nanodiamond 403 source, or size effect, we compared the EPR signal peak-to-404 peak line width (lwpp). It is worth noting that similar 18 nm-405 NDHPHT particles from a different provider have been 406 previously reported, 46 with a focus on particles of similar size 407 and characteristics.

Physicochemical Modifications of NDs. Air Oxidation 409 Process. ND_{DET} nanodiamonds have a complex surface nature 410 with a higher density of surface defects, amorphous and 411 graphitic carbons, and a wide diversity of functional groups, 412 which can be modulated by surface oxidation to generate acid 413 functions. Oxidation by air annealing provides an efficient 414 method for oxidizing ND surface and removing nondiamond 415 carbon, including paramagnetic radical-like centers (i.e., 416

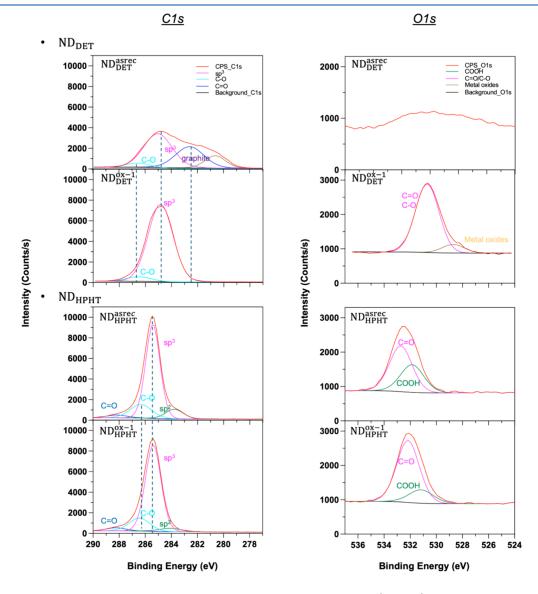


Figure 3. XPS data. Deconvoluted and experimental C 1s and O 1s fits of ND_{DET} and ND_{HPHT} (columns) obtained before and after air annealing for 4 h. Highlighted C 1s and O 1s data can be found in Table S3.

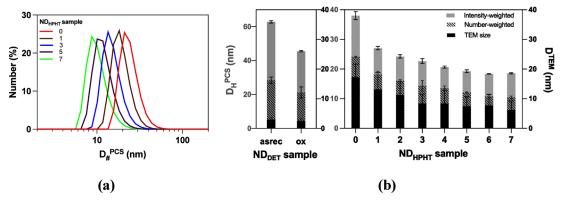


Figure 4. Size distributions. (a) Number size distributions obtained by size exclusion, starting from raw ND_{HPHT}^{assec} . Only ND_{HPHT} fragments isolated after one, three, five, and seven cycles (16,800g) of 60 min increment are shown for clarity. (b) Size distribution histograms by intensity- and number-weighted distributions and TEM sizes \pm standard deviations plotted as a function of the number of cycles (ND_{HPHT}^{assec} is written as "0", ND_{HPHT}^{1} referred to as "1", etc.) and comparison to ND_{DET} colloids. Note that the y axis ($D_{\#}^{PCS}$) for ND_{DET} and ND_{HPHT} samples is adjusted between nanodiamond sample types for clarity.

417 dangling bonds). Prior to this, thermogravimetric analyses 418 (TGAs) were performed on these materials to evaluate their 419 thermal behavior applying a heating rate of 10 °C/min (Figure 420 S1a); ND_{DET} and ND_{HPHT} begin to decompose at 515 and 483 421 °C, respectively, with a weight loss of 5%. A weight loss of 50% 422 is observed between 546 and 560 °C. Based on the information 423 resulting from TGA analysis, air-annealing conditions at 400 424 °C (ox-1), 450 °C (ox-2), 480 °C (ox-3), or 550 °C (ox-4) for 425 4 h were applied to evaluate the impact of oxidation on NDs' 426 physicochemical properties, especially with regard to their 427 surface chemical composition, graphitization, or dangling 428 bonds onto the ND surface. Thanks to the annealing process, 429 the weight decreases to 96 and 87% at 400 °C, 90 and 86% at 430 450 °C, 84 and 68% at 480 °C, and 31 and 1% at 550 °C after 431 4 h of air annealing for ND_{HPHT} and ND_{DET}, respectively 432 (Figure S2).

In ND_{DET} samples, primary particles appear as tightly 434 bounded clusters forming large aggregates in the suspension as 435 a result of their surface nature due to various functional groups 436 (i.e., ether, epoxide, hydroxyl, carbonyl, and carboxyl). The 437 first observation is that the applied treatments improve the 438 dispersion as well as the colloidal stability by preventing NDs 439 from aggregation. This observation is confirmed by PCS 440 analyses. While no significant difference in the core size was 441 noted by TEM (5.2 \pm 1.4 vs 4.5 \pm 1.2 nm), PCS indicates that 442 this treatment has a considerable decrease in the mean 443 hydrodynamic size from 63 nm ($D_{\#}^{PCS}$ = 28 nm) for ND_{DET} to 444 45-48 nm ($D_{\#}^{PCS}$ = 15-22 nm) for oxidized ND_{DET} samples, 445 respectively. The evolution of the zeta potential in relation to 446 pH changes indicates a shift from positive to negative values 447 (from $\zeta = +46$ to -44 mV after oxidation at pH 7.3; Figure 448 S3), suggesting an increase of colloidal stability through ionic 449 repulsion. Indeed, FTIR analysis reveals the emergence of a 450 carbonyl C=O stretching band around 1780 cm⁻¹, indicating 451 the formation of carboxylic groups on the surface. This is 452 responsible for the negative zeta potential, in concomitance 453 with the disappearance of C-H stretching modes that were 454 previously observed. Figure S4 also shows that the annealing at 455 400 °C leads to a spectrum similar to those obtained under 456 other conditions (up to 500 °C). In accordance with FTIR, 457 XPS data (Figure S5) showed an increase in the total oxygen 458 content in the ND_{DET} sample after air annealing (Table S3); 459 their atomic concentrations were calculated from the 460 corresponding C 1s and O 1s core-level peak intensities

(286-280 and 531-528 eV, respectively). The main differ- 461 ence in elemental composition in relative percentage terms is 462 expressed by the O 1s/C 1s ratio. While NDasrec nanodiamonds 463 contain 3.80% oxygen and 94.20% carbon (the remaining 2% 464 consists of nitrogen) (O 1s/C 1s = 0.04), ND_{DET}^{ox-1} nano- 465 diamonds contain 9.41% oxygen and 89.19% carbon (O 1s/C 466 1s = 0.10). This corresponds to a 59% increase in oxygen and a 467 5% reduction in carbon after air annealing under ox-1 468 conditions. Following the air-annealing treatment, the surface 469 composition of ND_{HPHT} seems to be less affected as suggested 470 by the slight evolution observed in the various FTIR spectra 471 (Figure S4). This observation suggests that, unlike ND_{DET}, the 472 ND_{HPHT}^{asrec} surface was already completely oxidized due to the 473 grinding process nature. 47 XPS (Figure 3) confirms those 474 f3 observations with no significant difference observed for 475 ND_{HPHT} samples under the same air-annealing procedure (O 476 1s/C 1s = 0.11). After their US-assisted dispersion in DI water, 477 PCS measurements indicate that controlled air annealing at 478 400, 450, 480, or 550 $^{\circ}$ C for 4 h compared to the ND $_{\mathrm{HPHT}}^{\mathrm{asrec}}$ 479 procedure has no influence on their colloidal behavior based 480 on their hydrodynamic particle size distribution profiles 481 (around $D_{\rm H}^{\rm PCS}$ 36 nm) and zeta potential values (around ζ 482 = -45 mV at pH 7.4). Overall, the quantitative analysis of the 483 evolution of the carboxylic acid content in ND_{DET} samples 484 indicated an increase of 19 to 27 nmol mg⁻¹ between ND_{DET} 485 and ND_{DET} conditions as determined by Boehm conducto- 486 metric titration. In the case of ND_{HPHT}, no significant 487 difference was noticed in these conditions (32 nmol mg⁻¹). 488 This average amount corresponds to 0.36 COOH groups per 489 nm² of the spherical ND_{DET} particle surface.

Size Exclusion Process. Due to the polydispersity of 491 ND_{HPHT}, we use a simple size exclusion approach to reduce 492 the particle size distribution and approximate the distribution 493 obtained for ND_{DET} after thermal treatment. The size sorting 494 process using centrifugation is a common and simple approach 495 to satisfying colloid fractionation. Previous reports showed 496 that MSY 0-0.05 (25 nm median size) is composed of 59% 497 (based on number-weight distribution) of NPs smaller than 17 498 nm, in particular, 33% of 5 nm ND_{HPHT}, as revealed by TEM 499 analysis. Herein, the high polydispersity (1.79) of the 500 ND_{HPHT} material offers the obvious advantage of being able 501 to be separable into distinct subdivision fractions starting from 502 the crude ND_{HPHT} (0.5%_{wt/v}). We assumed a similar 503 distribution in the MSY 0-0.03 sample. To roughly evaluate 504

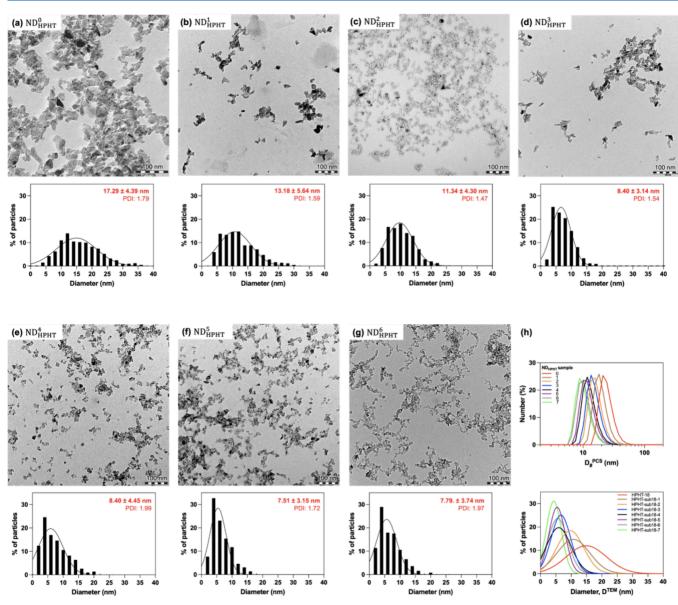


Figure 5. Size measurements on ND_{HPHT} size exclusion particles. TEM images (a–g) (scale bars: 100 nm) and statistical analysis after centrifugation with 60 min of increment starting from the ND_{HPHT}^{asrec} sample (core size approximately 18 nm). The histogram diameter axis is set by a 2 nm increment for clarity. (h) Dynamic size distributions (number-weighted distributions) of the isolated ND_{HPHT}^{ubb-17} fractions following centrifugation treatment at 16,800g starting from ND_{HPHT}^{asrec} (0 min), showing a shift toward smaller $D_{\#}^{PCS}$ distributions (up). Comparison of TEM size distributions using histograms fitted with a log-normal function (down).

505 the size distribution influenced by centrifugation, we employed 506 a constant high speed (16,800g) with extended durations in 60 507 min increments to generate a series of sub-17 nm fractions. 508 After each centrifugation cycle, the supernatant was carefully 509 separated to isolate the ND_{HPHT} sub-17 nm fractions. By 510 following this procedure, we were able to systematically reduce 511 the size of the ND_{HPHT} sample; substantial differences were 512 observed between the recorded distributions, indicating a shift 513 toward smaller particle diameters, as confirmed by PCS 514 measurements (Figure 4). Starting from the initial ND_{HPHT} 515 material ($D_{\#}^{PCS}$: 24.2 nm; PDI: 0.188), the number size 516 profiles clearly show a significant reduction in size distribution 517 well below $D_{\#}^{PCS}$ 13.2 nm (PDI: 0.164) after the first treatment 518 cycle (1 × 60 min; ND $_{HPHT}^{1}$ sample).

Efficient size exclusion was observed from the third s20 centrifugal treatment (ND_{HPHT}^3 sample; $D_{\#}^{PCS}$: 14.3 nm s21 (PDI: 0.132)). In correlation with DLS, TEM analysis

confirmed the PCS observations, showing an evolution of 522 the mean size from 17.3 ± 4.4 nm (PDI: 1.79) to 8.4 ± 3.1 nm 523 (PDI: 1.54) (Figure 5) for raw to ND $_{\rm HPHT}^3$ (37% in particle 524 f5 concentration), as depicted in the proposed sorting process. 525 The particle size of sub-17 nm HPHT thus approached that 526 obtained for detonation particles. No significant difference in 527 the size profile could be noticed when comparing ND $_{\rm HPHT}^3$ to 528 the other subfractions. Overall, this size treatment effectively 529 isolated the desired sub-17 nm particles, resulting in a relatively 530 monodisperse fraction (Table S4), despite a relatively low yield 531 (around $7.5\%_{\rm wt}$ in particle concentration), based on the initial 532 colloid concentration as obtained gravimetrically (Figure S6, 533 Table S4).

It must be mentioned that the separation of detonation 535 nanoparticles was not a straightforward applicable process; 536 indeed, 16,800g speed treatment led to a nonstable suspension 537 due to their aggregation states caused by the complex structure 538

Table 1. Comparison of the Physicochemical Parameters of Nanodiamonds

		size		EPR (X-band) $(g = 2.0027 \pm 0.0002)$		EPRI
sample	$\xi \text{ pH} = 7.3 \pm 0.1 \text{ (mV)}$	D _# PCS (nm)	$D_{\mathrm{H}}^{\mathrm{PCS}}$ (nm)	lwpp (mT)	N _S (spin/g)	R (mm)
ND _{DET}	+42 ± 2	28.4 ± 1.9	62.9 ± 0.7	0.96	6.6×10^{19}	>2
ND_{DET}^{ox-1}	-40 ± 2	21.2 ± 3.2	45.6 ± 0.4	0.88	5.8×10^{19}	>2
$\mathrm{ND}_{\mathrm{HPHT}}^{\mathrm{asrec}}$	-39 ± 2	24.2 ± 0.2	36.1 ± 0.2	0.38	1.5×10^{19}	0.7
ND^1_{HPHT}	-37 ± 2	27.1 ± 0.6	27.1 ± 0.6	0.27	1.4×10^{19}	>0.5
ND_{HPHT}^{3}	-36 ± 1	14.3 ± 1.8	23.9 ± 0.3	0.23	1.3×10^{19}	0.5
ND_{HPHT}^5	-44 ± 2	12.2 ± 0.2	19.3 ± 0.4	0.23	1.0×10^{19}	< 0.5
ND_{HPHT}^{7}	-44 ± 2	10.4 ± 0.5	18.6 ± 0.2	0.22	9.8×10^{18}	< 0.5
ND_{HPHT}^{ox-1}	-43 ± 1	22.6 ± 0.6	35.4 ± 0.1	0.28	1.6×10^{19}	>0.5
ND_{HPHT}^{ox-2}	-46 ± 1	21.2 ± 0.8	34.5 ± 0.2	0.27	9.1×10^{18}	>0.5
ND_{HPHT}^{ox-3}	-46 ± 1	21.0 ± 1.2	37.8 ± 0.1	0.26	6.7×10^{18}	0.5
ND_{HPHT}^{ox-4}	-47 ± 1	15.6 ± 1.1	32.6 ± 0.5	0.24	2.1×10^{18}	<0.5

539 of the isolated detonation shoot. Photos of as-received and 540 thermally oxidized particles treated for 60 min at 16,800g are 541 shown in Figure S7. This procedure was not applied to the 542 detonation nanodiamonds due to a lack of colloidal stability 543 resulting from their aggregation state. We note that these 544 particles are usually tightly bound into aggregates despite many 545 efforts (milling aggregates with micrometer-sized ceramic 546 beads) to stabilize individual spherical NPs. 48

EPR Spectroscopic and Imaging Study for Spatial 548 Resolution. HPHT-Synthesized Nanodiamonds. Interest-549 ingly, a noticeable line width reduction effect can be observed 550 in EPR spectroscopy from 0.38 mT onward as the average 551 particle size decreases through size exclusion. The EPR 552 features (signal width) of this series of HPHT are presented 553 in Table 1. The line width decreased and then reached a 554 plateau from sample ND_{HPHT}^3 to ND_{HPHT}^5 (ND_{HPHT}^3 ; $N_s = 1.3$ $555 \times 10^{19} \text{ spin/g}$) for a value of around 0.23 mT, corresponding 556 to a decrease in the broad component of nearly 40%, for which 557 the ND_{HPHT} sample appeared as an efficient sample for EPRI 558 investigation (achievable resolution < 0.5 nm). This set of size 559 variations in ND_{HPHT} resulted in a weak decrease of spins per 560 gram (N_S) but, strikingly, an efficient decrease in the broad 561 component of the signal. The comparison of the physicochem-562 ical parameters collected for the different series of nano-563 diamonds is summarized in Table 1.

Alternatively, thermal annealing of ND_{HPHT} resulted in a sess decrease in the broad component of the signal. In contrast to detonation nanodiamonds, the air-annealing process has a minimal impact on the spins located on the outer surface due to the surface nature of HPHT particles.

Although FTIR spectroscopy did not reveal any significant 570 qualitative differences between the samples after thermal 571 treatments, there was a significant impact on their lwpp. 572 Raman spectroscopy was used to determine these differences. 573 This technique provided additional information about their 574 structural changes. The Raman spectrum of ND_{HPHT} is shown 575 in Figure S8a and displays the following characteristics: ⁴⁹ (i) a 576 peak at 1330 cm⁻¹ (dotted line) associated with the first-order 577 diamond peak (C*, diamond phase); (ii) a deconvoluted peak 578 at 1350 cm⁻¹ assigned to sp²-hybridized bond deformation 579 mode (D-band); (iii) 1470 cm⁻¹ that could be attributed to 580 the presence of nitrogen-vacancy defects in the diamond 581 lattice; (iv) a broad component at 1570-1590 cm⁻¹ due to 582 allotropic forms of carbon sp²-graphitic carbon covering the 583 diamond core (G-band); and (v) a band at 1685 cm⁻¹ 584 characteristic of C-OH/C=O bending vibrations at the 585 particle surface. In addition to the above-mentioned peaks, the

ND_{HPHT} 1580 cm⁻¹ peak (G-band) decreases due to the air- 586 annealing progress of HPHT samples (400 to 550 °C) among 587 the other nondiamonds' absorption (Figure S8a). Indeed, 588 Raman spectra of HPHT particles show a considerable 589 increase in the diamond peak (C*) intensity compared to 590 the broad *sp*²-related band (1500–1800 cm⁻¹) of the air- 591 annealed samples, and its shift 1332–1333 cm⁻¹ becomes 592 dominant (Figure S8b). It is evident that the air-annealing of 593 the HPHT powder at 550 °C for 4 h (ND_{HPHT}) considerably 594 reduces the graphitic content as the G-band is further 595 suppressed.

For both strategies, these distinct treatments enabled 597 adequate control of the size and surface area, leading to the 598 fine-tuning of parameters and lower $N_{\rm S}$ content. Overall, the 599 EPR signal of HPHT particles typically exhibits a narrower line 600 width for the narrow component compared to other types of 601 nanodiamonds. This suggests a higher degree of order in the 602 material, although line widths are also influenced by other 603 factors, such as the initial content of P1 centers and exchange 604 interactions between spins within the particle. This is 605 particularly true as the size decreases from micrometers (100 606 μ m) to nanometers (20 nm) as reported in the literature. 608 appeared in the EPR spectrum when the average size of 609 particles in polydisperse samples decreases. 610

DET Nanodiamonds. In contrast, the EPR signal of 611 detonation nanodiamonds typically exhibits a broad line 612 width (lwpp = 0.96 mT) due to the presence of multiple 613paramagnetic defects such as structural defects, dangling 614 bonds, impurities, and graphite and diamond allotropes. 51 615 Overall, HPHT nanodiamonds exhibit a narrower signal than 616 NDasrec, representing a reduction of almost 60% of the fwhm 617 (full width at half-maximum). In addition, the concentration of 618 RPC (spin S = 1/2) was estimated to be $N_s = 1.5 \times 10^{19}$ spin/g 619 on the HPHT colloid, which is ~4.4 times less than the 620 detonation sample ($N_s = 6.6 \times 10^{19} \text{ spin/g}$). Assuming that 621 NDasrec particles are spherical, an experimental value of 622 approximately 5 RPC/particle is obtained, and the spins are 623 inhomogeneously distributed according to the literature. 48,51 624 Interestingly, we observe a reduction in the EPR signal line 625 width resulting from the decrease in spin density at the particle 626 surface during air-oxidation treatment. For comparison, 627 oxidative treatment of ND_{DET} reduces the broad component 628 by 10%, associated with impurities near the particle surface 629 (mainly dangling bonds) for the ox-1 sample. This fact 630 illustrates the importance of surface properties and sets for 631 surface development.

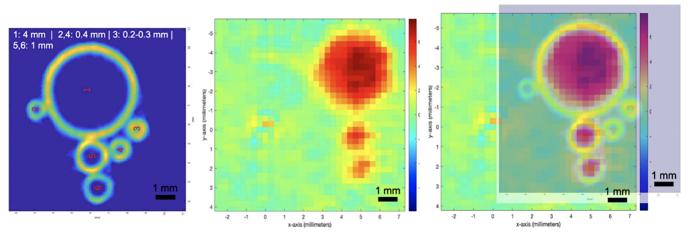


Figure 6. L-band EPR imaging experiment of the ND_{ASPEC} sample. Anatomic micro-CT image (left), corresponding 2D image (ZY) of glass capillaries containing ND suspensions imaged by EPRI (middle), and the superimposed spatial registration of the EPR image to the corresponding micro-CT scan (right). EPRI parameters: experimental time: 71 min; lwpp: 0.35 mT; and FOV: 37.55 mm.

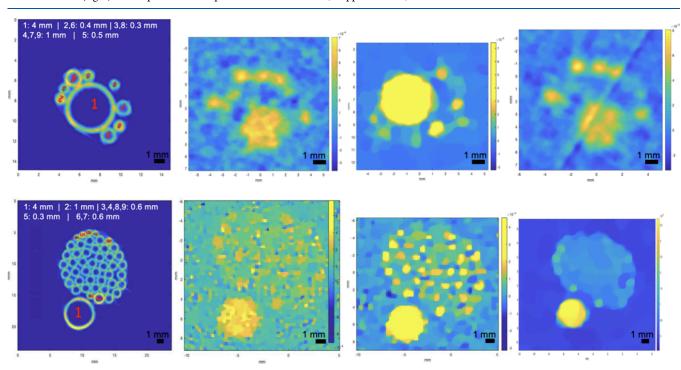


Figure 7. L-band EPR images of the ND $_{\rm HPHT}^3$ sample. EPRI and the phantom object anatomic CT-microtomography (left) and 2D images (ZY) of glass capillaries containing aqueous suspensions of NDs from left to right according to iteration number, regularity parameter λ' . ND densities of 10 mg mL $^{-1}$ (up) and 20 mg mL $^{-1}$ (down) were imaged.

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The main advantage of ND_{HPHT} over ND_{DET} is the uniform 634 crystallinity and purity with a relatively low and localized concentration of spin-resonant lattice defects. This is crucial 636 for their spectroscopic characteristics and imaging applications, as they can stably host narrow single-line EPR components. The concentration of radical-like paramagnetic centers, 638 639 including impurities and dangling bonds, can be well-suited 640 to EPRI applications with significant decreases in peak-to-peak 641 derived line width after ND_{HPHT} size exclusion. Three 642 centrifugation cycles of the ND_{HPHT} nanodiamonds yield 643 promising EPR features (minimal lwpp obtained = 0.22 mT). 644 In an interesting feature involving nanodiamonds, we highlight 645 how the use of NDs can be relevant to afford a unique EPR 646 contrast signature through long-term stable paramagnetic 647 centers.

L-Band EPR Imaging Study for Spatial Resolution. 648 Imaging capabilities of NDs have been further evaluated in L- 649 band (1 GHz) EPR spectroscopy (Figure S9) and imaging for 650 low-field frequency measurements. EPR images are recorded 651 similarly to a conventional EPR spectrum, with magnetic field 652 gradients applied in a range of different orientations around the 653 sample (gradient = 4 mT/cm). The achievable resolution in an 654 EPRI experiment is based on concepts similar to those of MRI 655 acquisition. Formally, a common definition of resolution is the 656 determination of the number of pixels in a specified field of 657 view (FOV). In this case, the theoretical spatial resolution in 658 EPRI is closely proportional to the peak-to-peak line width of 659 the EPR spectroscopic absorption curve (lwpp) and the 660 inverse of the gradient intensity (∇); typically, $R = \text{lwpp}/\nabla$. 661 Overall, a narrower line width and stronger gradients allow for 662

high-resolution EPR images ($\lim_{V\to\infty} R=0$), which are limited by 664 the magnetic field produced by the gradients⁵² for small 665 gradient strength (∇ < 10% principal magnetic field). Two-666 dimensional imaging also implies spatial resolution represented 667 by the pixel resolution (R_{Pixel} = FOV/number of pixel) of the 668 imaging system with correct image reconstruction. 41 However, 669 broad peak-to-peak line widths can limit the application of 670 various types of NPs for resolution enhanced imaging, which 671 tends to give special attention to nanometric ND_{HPHT} . In this 672 case, by employing a deconvolution process to recover a 673 spatially homogeneous spin density distribution, spatial 674 resolutions of less than 0.7 mm could be obtained in the 675 improved EPR imaging system starting from the raw ND_{HPHT} 676 sample. Using mathematical algorithms and imaging opti-677 mizations, we present promising preliminary EPRI acquisitions 678 of capillary phantoms after image reconstruction. The 679 experimental resolution of ND_{HPHT} was evaluated by image 680 analysis based on quartz capillaries containing mean 18 nm size 681 monocrystalline particles with inner-diameter sizes of 0.2, 0.3, 682 0.4, 0.5, 0.6, 1, and 4.0 mm. Imaging of raw ND_{HPHT}^{asrec} (lwpp = 683 0.38 mT; RPC = 0.442 mM) filled into capillaries at a similar 684 concentration (10 mg mL⁻¹) showed a strong detectable 685 signal. An achievable resolution of <0.7 mm was observed on 686 Figure 6 with spatial registration and fusion of raw EPRI and 687 X-ray micro-CT on the same object. In contrast, ND_{DET} (lwpp 688 = 0.98 mT) did not provide the expected resolution since R > 100 m689 2 mm under the data acquisitions (R = 0.23 mm for a FOV of 690 32 mm) (data not shown). We can mention that the ND_{DET} 691 EPR signal operating at microwave frequency of 1 GHz 692 presents a known spectral resolution limitation compared to 693 the ND_{HPHT} sample. Despite a 4.4-fold decrease in RPC

Additionally, as previously mentioned, the spatial resolution of EPR images (R) depends on probe line widths and the 700 magnetic field strength. Thus, reducing the line width of the 701 sample may enhance image contrast. From all the samples 702 under study, we consider the ND_{HPHT} colloid to be the most 703 promising (lwpp = 0.23 mT). Furthermore, if the number of 704 spins in a pixel or voxel falls below the detection threshold 705 (<0.4 mm), no image can be acquired under these conditions. 706 On this basis, we investigate the nanodiamond density of the 707 ND $_{\rm HPHT}^3$ sample by studying a 10 mg mL $^{-1}$ (RPC = 0.221 708 mM) (Figure 7; upper line) or 20 mg mL $^{-1}$ (RPC = 0.442 709 mM) (Figure 7; lower line) suspension. Full details specific to 710 sampling imaging parameters can be found in the paper from 711 Abergel et al. 42 Images were processed according to the 712 number of iterations.

694 content compared with $\ensuremath{\text{ND}_{\text{DET}}}\xspace$, typically exhibiting a spin

695 density $N_{\rm S}$ of 6.6 \times 10¹⁹ vs 1.5 \times 10¹⁹ spin/g for ND_{DET} and

 696 ND $^{asrec}_{HPHT}$, respectively, the latter still demonstrates good

sensitivity and threshold detection.

The images above represent the first combined EPRI and 714 micro-CT images offering good resolution of a phantom (R < 715 0.5 mm) and open the way to the utilization of NDs in the 716 field of EPR imaging.

717 CONCLUSIONS

718 This study explores the physicochemical properties of 719 nanodiamonds, focusing on size and surface treatments using 720 commercial materials obtained by grinding or detonation 721 synthesis. The difference in production conditions allows one 722 to differ the chemical composition of impurities and their 723 distribution along the crystal as well as control over a

predetermined nanoparticle size and shape. Using size 724 exclusion on a polydisperse HPHT sample (mean size 18 725 nm), a particle size distribution similar to that of detonation 726 nanodiamonds was achieved with improved structural quality 727 and surface uniformity. In addition, we describe thermal 728 oxidation because it is a straightforward approach that allows 729 surface chemistry uniformization leading to oxygen-terminated 730 particles and comparison of EPR features. Furthermore, EPR 731 spectroscopy revealed the promising properties of HPHT 732 nanodiamonds for probing at smaller sizes, demonstrating the 733 capabilities of low-field (1 GHz) EPR imaging. Overall, this 734 study highlights the unique EPR properties of nanodiamonds, 735 specifically the ND_{HPHT} sample (mean size of 8 nm), and offers 736 prospects for applications in nanomedicine and materials 737 science.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at 741 https://pubs.acs.org/doi/10.1021/acs.langmuir.4c05169.

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HRTEM-EDX data (Table S1), ICP data (Table S2), 743 XPS data (Table S3), size data (Table S4), surface 744 analysis (Figure S1), gravimetric measurement (Figure 745 S2), zeta potential data (Figure S3), IR (Figure S4), XPS 746 spectra (Figure S4), size obtained by size exclusion 747 (Figure S6), photographs of ND suspensions through 748 size exclusion process (Figure S7), Raman spectra 749 (Figure S8), and L-band EPR spectrum of ND sample 750 (Figure S9) (PDF)

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793 Notes

794 The authors declare no competing financial interest.

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809 REFERENCES

- 810 (1) Danhier, P.; Gallez, B. Electron Paramagnetic Resonance: A 811 Powerful Tool to Support Magnetic Resonance Imaging Research. 812 Contrast Media Mol. Imaging 2015, 10 (4), 266–281.
- 813 (2) Kempe, S.; Metz, H.; Mäder, K. Application of Electron 814 Paramagnetic Resonance (EPR) Spectroscopy and Imaging in Drug 815 Delivery Research Chances and Challenges. *Eur. J. Pharm.* 816 *Biopharm.* 2010, 74 (1), 55–66.
- 817 (3) Abbas, K.; Babić, N.; Peyrot, F. Use of Spin Traps to Detect 818 Superoxide Production in Living Cells by Electron Paramagnetic 819 Resonance (EPR) Spectroscopy. *Methods* **2016**, *109*, 31–43.
- 820 (4) Bordignon, E. EPR Spectroscopy of Nitroxide Spin Probes. In 821 *eMagRes*; Harris, R. K.; Wasylishen, R. L., Eds.; John Wiley & Sons, 822 Ltd: Chichester, UK, 2017; pp 235–254.
- 823 (5) Tran, D.-N.; Li-Thiao-Te, S.; Frapart, Y.-M. Parameter 824 Estimation for Quantitative EPR Spectroscopy. *IEEE Trans. Instrum.* 825 *Meas.* **2021**, *70*, 1–7.
- 826 (6) Krishna, M. C.; Subramanian, S. The Development of Time-827 Domain In Vivo EPR Imaging at NCI. *Appl. Magn. Reson.* **2021**, 52 828 (10), 1291–1309.
- 829 (7) Sahu, I. D.; Lorigan, G. A. Electron Paramagnetic Resonance as a 830 Tool for Studying Membrane Proteins. *Biomolecules* **2020**, *10* (5), 831 763.
- 832 (8) Hofmann, L.; Ruthstein, S. EPR Spectroscopy Provides New 833 Insights into Complex Biological Reaction Mechanisms. *J. Phys. Chem.* 834 B **2022**, 126 (39), 7486–7494.
- 835 (9) Galazzo, L.; Bordignon, E. Electron Paramagnetic Resonance 836 Spectroscopy in Structural-Dynamic Studies of Large Protein 837 Complexes. *Prog. Nucl. Magn. Reson. Spectrosc.* **2023**, 134–135, 1–19. 838 (10) Passeri, D.; Rinaldi, F.; Ingallina, C.; Carafa, M.; Rossi, M.;
- 839 Terranova, M. L.; Marianecci, C. Biomedical Applications of 840 Nanodiamonds: An Overview. *J. Nanosci. Nanotechnol.* **2015**, 15 841 (2), 972–988.
- 842 (11) Chauhan, S.; Jain, N.; Nagaich, U. Nanodiamonds with 843 Powerful Ability for Drug Delivery and Biomedical Applications:

- Recent Updates on in Vivo Study and Patents. J. Pharm. Anal. 2020, 844 10 (1), 1–12.
- (12) Garifo, S.; Stanicki, D.; Ayata, G.; Muller, R. N.; Laurent, S. 846 Nanodiamonds as Nanomaterial for Biomedical Field. *Front. Mater.* 847 *Sci.* 2021, 15 (3), 334–351.
- (13) Pichot, V.; Muller, O.; Seve, A.; Yvon, A.; Merlat, L.; Spitzer, D. 849 Optical Properties of Functionalized Nanodiamonds. *Sci. Rep.* **2017**, 7 850 (1), 14086.
- (14) Ekimov, E. A.; Shiryaev, A. A.; Grigoriev, Y.; Averin, A.; 852 Shagieva, E.; Stehlik, S.; Kondrin, M. V. Size-Dependent Thermal 853 Stability and Optical Properties of Ultra-Small Nanodiamonds 854 Synthesized under High Pressure. *Nanomater.* 2022, 12 (3), 351.
- (15) Turcheniuk, K.; Mochalin, V. N. Biomedical Applications of 856 Nanodiamond (Review). Nanotechnol. 2017, 28 (25), 252001.
- (16) Ansari, S. A.; Satar, R.; Jafri, M. A.; Rasool, M.; Ahmad, W.; 858 Kashif Zaidi, S. Role of Nanodiamonds in Drug Delivery and Stem 859 Cell Therapy. *Iran. J. Biotechnol.* **2016**, *14* (3), 130–141.
- (17) Krok, E.; Balakin, S.; Jung, J.; Gross, F.; Opitz, J.; Cuniberti, G. 861 Modification of Titanium Implants Using Biofunctional Nano- 862 diamonds for Enhanced Antimicrobial Properties. *Nanotechnol.* 863 **2020**, 31 (20), 205603.
- (18) Singh, D.; Ray, S. A Short Appraisal of Nanodiamonds in Drug 865 Delivery and Targeting: Recent Advancements. *Front. Nanotechnol.* 866 **2023**, *5*, 1259648.
- (19) Wu, Y.; Alam, M. N. A.; Balasubramanian, P.; Ermakova, A.; 868 Fischer, S.; Barth, H.; Wagner, M.; Raabe, M.; Jelezko, F.; Weil, T. 869 Nanodiamond Theranostic for Light-Controlled Intracellular Heating 870 and Nanoscale Temperature Sensing. *Nano Lett.* **2021**, 21 (9), 3780—871 3788.
- (20) Panich, A. M.; Salti, M.; Goren, S. D.; Yudina, E. B.; 873 Aleksenskii, A. E.; Vul, A. Y.; Shames, A. I. Gd(III)-Grafted 874 Detonation Nanodiamonds for MRI Contrast Enhancement. *J. Phys.* 875 Chem. C 2019, 123 (4), 2627–2631.
- (21) Panich, A. M.; Salti, M.; Prager, O.; Swissa, E.; Kulvelis, Y. V.; 877 Yudina, E. B.; Aleksenskii, A. E.; Goren, S. D.; Vul, A. Ya.; Shames, A. 878 I. PVP-coated Gd-grafted Nanodiamonds as a Novel and Potentially 879 Safer Contrast Agent for in Vivo MRI. *Magn. Reson. Med.* **2021**, 86 880 (2), 935–942.
- (22) Kunuku, S.; Lin, B.-R.; Chen, C.-H.; Chang, C.-H.; Chen, T.-Y.; 882 Hsiao, T.-Y.; Yu, H.-K.; Chang, Y.-J.; Liao, L.-C.; Chen, F.-H.; 883 Bogdanowicz, R.; Niu, H. Nanodiamonds Doped with Manganese for 884 Applications in Magnetic Resonance Imaging. ACS Omega 2023, 8 885 (4), 4398–4409.
- (23) Panich, A. M.; Shames, A. I.; Aleksenskii, A. E.; Yudina, E. B.; 887 Vul, A. Y. Manganese-Grafted Detonation Nanodiamond, a Novel 888 Potential MRI Contrast Agent. *Diamond Relat. Mater.* **2021**, *119*, 889 No. 108590.
- (24) Panich, A. M.; Salti, M.; Aleksenskii, A. E.; Kulvelis, Y. V.; 891 Chizhikova, A.; Vul, A. Ya.; Shames, A. I. Suspensions of Manganese- 892 Grafted Nanodiamonds: Preparation, NMR, and MRI Study. *Diam.* 893 *Relat. Mater.* **2023**, *131*, No. 109591.
- (25) Garifo, S.; Vangijzegem, T.; Stanicki, D.; Laurent, S. A Review 895 on the Design of Carbon-Based Nanomaterials as MRI Contrast 896 Agents. *Mol.* **2024**, *29* (7), 1639.
- (26) Waddington, D. E. J.; Boele, T.; Rej, E.; McCamey, D. R.; King, 898 N. J. C.; Gaebel, T.; Reilly, D. J. Phase-Encoded Hyperpolarized 899 Nanodiamond for Magnetic Resonance Imaging. *Sci. Rep.* **2019**, 9 (1), 900 5950.
- (27) Waddington, D. E. J.; Sarracanie, M.; Zhang, H.; Salameh, N.; 902 Glenn, D. R.; Rej, E.; Gaebel, T.; Boele, T.; Walsworth, R. L.; Reilly, 903 D. J.; Rosen, M. S. Nanodiamond-Enhanced MRI via in Situ 904 Hyperpolarization. *Nat. Commun.* **2017**, *8* (1), 15118.
- (28) Ajoy, A.; Nazaryan, R.; Druga, E.; Liu, K.; Aguilar, A.; Han, B.; 906 Gierth, M.; Oon, J. T.; Safvati, B.; Tsang, R.; Walton, J. H.; Suter, D.; 907 Meriles, C. A.; Reimer, J. A.; Pines, A. Room Temperature "Optical 908 Nanodiamond Hyperpolarizer": Physics, Design, and Operation. Rev. 909 Sci. Instrum. 2020, 91 (2), No. 023106.
- (29) Bobko, A. A.; Kirilyuk, I. A.; Grigor'ev, I. A.; Zweier, J. L.; 911 Khramtsov, V. V. Reversible Reduction of Nitroxides to Hydroxyl- 912

- 913 amines: Roles for Ascorbate and Glutathione. Free Radic. Biol. Med. 914 2007, 42 (3), 404–412.
- 915 (30) Jaymand, M.; Davatgaran Taghipour, Y.; Rezaei, A.; 916 Derakhshankhah, H.; Foad Abazari, M.; Samadian, H.; Hamblin, M.
- 917 R. Radiolabeled Carbon-Based Nanostructures: New Radiopharma-918 ceuticals for Cancer Therapy? *Coord. Chem. Rev.* **2021**, 440, 919 No. 213974.
- 920 (31) Mochalin, V. N.; Shenderova, O.; Ho, D.; Gogotsi, Y. The 921 Properties and Applications of Nanodiamonds. *Nat. Nanotechnol.* 922 **2012**, 7 (1), 11–23.
- 923 (32) Qin, J.-X.; Yang, X.-G.; Lv, C.-F.; Li, Y.-Z.; Liu, K.-K.; Zang, J.-924 H.; Yang, X.; Dong, L.; Shan, C.-X. Nanodiamonds: Synthesis, 925 Properties, and Applications in Nanomedicine. *Mater. Des.* **2021**, *210*,
- 927 (33) Whitlow, J.; Pacelli, S.; Paul, A. Multifunctional Nanodiamonds 928 in Regenerative Medicine: Recent Advances and Future Directions. 929 JCR 2017, 261, 62–86.
- 930 (34) Kovářík, T.; Bělský, P.; Rieger, D.; Ilavsky, J.; Jandová, V.; 931 Maas, M.; Šutta, P.; Pola, M.; Medlín, R. Particle Size Analysis and 932 Characterization of Nanodiamond Dispersions in Water and 933 Dimethylformamide by Various Scattering and Diffraction Methods. 934 J. Nanopart. Res. 2020, 22 (2), 34.
- 935 (35) Stehlik, S.; Varga, M.; Ledinsky, M.; Jirasek, V.; Artemenko, A.; 936 Kozak, H.; Ondic, L.; Skakalova, V.; Argentero, G.; Pennycook, T.; 937 Meyer, J. C.; Fejfar, A.; Kromka, A.; Rezek, B. Size and Purity Control 938 of HPHT Nanodiamonds down to 1 Nm. *J. Phys. Chem. C* 2015, 119 939 (49), 27708–27720.
- 940 (36) Rehor, I.; Cigler, P. Precise Estimation of HPHT Nano-941 diamond Size Distribution Based on Transmission Electron 942 Microscopy Image Analysis. *Diam. Relat. Mater.* **2014**, *46*, 21–24.
- 943 (37) Hannecart, A.; Stanicki, D.; Vander Elst, L.; Muller, R. N.; 944 Brûlet, A.; Sandre, O.; Schatz, C.; Lecommandoux, S.; Laurent, S. 945 Embedding of Superparamagnetic Iron Oxide Nanoparticles into 946 Membranes of Well-Defined Poly(Ethylene Oxide)- *Block* -Poly(*e*-947 Caprolactone) Nanoscale Magnetovesicles as Ultrasensitive MRI 948 Probes of Membrane Bio-Degradation. *J. Mater. Chem. B* **2019**, 7 949 (30), 4692–4705.
- 950 (38) Schmidlin, L.; Pichot, V.; Comet, M.; Josset, S.; Rabu, P.; 951 Spitzer, D. Identification, Quantification and Modification of 952 Detonation Nanodiamond Functional Groups. *Diam. Relat. Mater.* 953 **2012**, 22, 113–117.
- 954 (39) Volkov, D. S.; Proskurnin, M. A.; Korobov, M. V. Elemental 955 Analysis of Nanodiamonds by Inductively-Coupled Plasma Atomic 956 Emission Spectroscopy. *Carbon* **2014**, *74*, 1–13.
- 957 (40) Stoll, S.; Schweiger, A. EasySpin, a Comprehensive Software 958 Package for Spectral Simulation and Analysis in EPR. *J. Magn. Reson.* 959 **2006**, 178 (1), 42–55.
- 960 (41) Durand, S.; Frapart, Y.-M.; Kerebel, M. Electron Paramagnetic 961 Resonance Image Reconstruction with Total Variation and Curvelets 962 Regularization. *Inverse Probl.* **2017**, 33 (11), 114002.
- 963 (42) Abergel, R.; Boussâa, M.; Durand, S.; Frapart, Y.-M. Electron 964 Paramagnetic Resonance Image Reconstruction with Total Variation 965 Regularization. *IPOL* **2023**, *13*, 90–139.
- 966 (43) Stehlik, S.; Mermoux, M.; Schummer, B.; Vanek, O.; Kolarova, 967 K.; Stenclova, P.; Vlk, A.; Ledinsky, M.; Pfeifer, R.; Romanyuk, O.; 968 Gordeev, I.; Roussel-Dherbey, F.; Nemeckova, Z.; Henych, J.; 969 Bezdicka, P.; Kromka, A.; Rezek, B. Size Effects on Surface Chemistry 970 and Raman Spectra of Sub-5 Nm Oxidized High-Pressure High-971 Temperature and Detonation Nanodiamonds. *J. Phys. Chem. C* 2021, 972 125 (10), 5647–5669.
- 973 (44) Dolmatov, V. Yu.; Lapchuk, N. M.; Lapchuk, T. M.; Nguyen, B. 974 T. T.; Myllymäki, V.; Vehanen, A.; Yakovlev, R. Yu. A Study of 975 Defects and Impurities in Doped Detonation Nanodiamonds by EPR, 976 Raman Scattering, and XRD Methods. *J. Superhard Mater.* **2016**, 38

977 (4), 219-229.

978 (45) Shames, A. I.; Zegrya, G. G.; Samosvat, D. M.; Osipov, V. Y.; 979 Vul, A. Y. Size Effect in Electron Paramagnetic Resonance Spectra of 980 Impurity Centers in Diamond Particles. *Phys. E* **2023**, *146*, 981 No. 115523.

- (46) Panich, A. M.; Sergeev, N. A.; Shames, A. I.; Osipov, V. Y.; 982 Boudou, J.-P.; Goren, S. D. Size Dependence of C Nuclear Spin-983 Lattice Relaxation in Micro- and Nanodiamonds. *J. Phys.: Condens.* 984 *Matter.* **2015**, 27 (7), No. 072203.
- (47) Petit, T.; Puskar, L. FTIR Spectroscopy of Nanodiamonds: 986 Methods and Interpretation. *Diam. Relat. Mater.* **2018**, 89, 52–66. 987 (48) Panich, A. M. Nuclear Magnetic Resonance Studies of 988 Nanodiamonds. *Crit. Rev. Solid State Mater. Sci.* **2012**, 37 (4), 276–989 303.
- (49) Mermoux, M.; Chang, S.; Girard, H. A.; Arnault, J.-C. Raman 991 Spectroscopy Study of Detonation Nanodiamond. *Diam. Relat. Mater.* 992 **2018**, 87, 248–260. 993
- (50) Zegrya, G. G.; Samosvat, D. M.; Osipov, V. Y.; Vul, A. Y.; 994 Shames, A. I. Size Effect in Electron Paramagnetic Resonance Spectra 995 of Impurity Centers in Diamond Nanoparticles. *Mesoscale Nanoscale Phys.* **2019**, 1–25.
- (51) Shames, A. I.; Panich, A. M.; Kempiński, W.; Alexenskii, A. E.; 998 Baidakova, M. V.; Dideikin, A. T.; Osipov, V. Y.; Siklitski, V. I.; 999 Osawa, E.; Ozawa, M.; Vul, A. Y. Defects and Impurities in 1000 Nanodiamonds: EPR, NMR and TEM Study. J. Phys. Chem. Solids 1001 2002, 63 (11), 1993–2001.
- (52) Bézière, N.; Decroos, C.; Mkhitaryan, K.; Kish, E.; Richard, F.; 1003 Bigot-Marchand, S.; Durand, S.; Cloppet, F.; Chauvet, C.; Corvol, M.- 1004 T.; Rannou, F.; Xu-Li, Y.; Mansuy, D.; Peyrot, F.; Frapart, Y.-M. First 1005 Combined in Vivo X-Ray Tomography and High-Resolution 1006 Molecular Electron Paramagnetic Resonance (EPR) Imaging of the 1007 Mouse Knee Joint Taking into Account the Disappearance Kinetics of 1008 the EPR Probe. *Mol. Imaging* 2012, 11 (3), 220.